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| 09/973,199 10/10/2001 | | 10/10/2001 | Bangalore Eahwar Amita Rani | 056859-0131 4508 | |
| 22428 | 7590 | 11/02/2006 | | EXAMINER | |
| FOLEY AN | ID LARI | ONER LLP | HUYNH, PHUONG N | | |
| SUITE 500 3000 K STR | EET NW | | ART UNIT | PAPER NUMBER | |
| WASHINGT | ON, DC | 20007 | 1644 | | |

DATE MAILED: 11/02/2006

Please find below and/or attached an Office communication concerning this application or proceeding.

| | | Application No. | Applicant(s) | | | | | |
|--|---|---|-------------------|--------|--|--|--|--|
| | Office Addison October | 09/973,199 | RANI ET AL. | | | | | |
| | Office Action Summary | Examiner | Art Unit | | | | | |
| | | Phuong Huynh | 1644 | | | | | |
| Period fo | The MAILING DATE of this communication app or Reply | ears on the cover sheet with the d | correspondence ad | ldress | | | | |
| A SHORTENED STATUTORY PERIOD FOR REPLY IS SET TO EXPIRE three MONTH(S) OR THIRTY (30) DAYS, WHICHEVER IS LONGER, FROM THE MAILING DATE OF THIS COMMUNICATION. - Extensions of time may be available under the provisions of 37 CFR 1.136(a). In no event, however, may a reply be timely filed after SIX (6) MONTHS from the mailing date of this communication. - If NO period for reply is specified above, the maximum statutory period will apply and will expire SIX (6) MONTHS from the mailing date of this communication. - Failure to reply within the set or extended period for reply will, by statute, cause the application to become ABANDONED (35 U.S.C. § 133). Any reply received by the Office later than three months after the mailing date of this communication, even if timely filed, may reduce any earned patent term adjustment. See 37 CFR 1.704(b). | | | | | | | | |
| Status | · | | | | | | | |
| 1) 又 | Responsive to communication(s) filed on 11 Au | aust 2006. | | - • | | | | |
| · | · · · · · · · · · · · · · · · · · · · | action is non-final. | | | | | | |
| ′ | , — | Since this application is in condition for allowance except for formal matters, prosecution as to the merits is | | | | | | |
| ,— | closed in accordance with the practice under <i>Ex parte Quayle</i> , 1935 C.D. 11, 453 O.G. 213. | | | | | | | |
| Dispositi | on of Claims | | | | | | | |
| 4)⊠ | 4)⊠ Claim(s) <u>1 and 5-15</u> is/are pending in the application. | | | | | | | |
| •— | 4a) Of the above claim(s) is/are withdrawn from consideration. | | | | | | | |
| | ☐ Claim(s) is/are allowed. | | | | | | | |
| 6)⊠ | Claim(s) <u>1 and 5-15</u> is/are rejected. | | | | | | | |
| 7) | | | | | | | | |
| 8)□ | Claim(s) are subject to restriction and/or election requirement. | | | | | | | |
| Applicati | on Papers | | | | | | | |
| 9) | The specification is objected to by the Examine | r. | | | | | | |
| 10) The drawing(s) filed on is/are: a) accepted or b) objected to by the Examiner. | | | | | | | | |
| Applicant may not request that any objection to the drawing(s) be held in abeyance. See 37 CFR 1.85(a). | | | | | | | | |
| Replacement drawing sheet(s) including the correction is required if the drawing(s) is objected to. See 37 CFR 1.121(d). | | | | | | | | |
| 11) The oath or declaration is objected to by the Examiner. Note the attached Office Action or form PTO-152. | | | | | | | | |
| Priority u | ınder 35 U.S.C. § 119 | | | | | | | |
| 12) Acknowledgment is made of a claim for foreign priority under 35 U.S.C. § 119(a)-(d) or (f). a) All b) Some * c) None of: | | | | | | | | |
| | 1. Certified copies of the priority documents have been received. | | | | | | | |
| | 2. Certified copies of the priority documents have been received in Application No | | | | | | | |
| 3. Copies of the certified copies of the priority documents have been received in this National Stage | | | | | | | | |
| application from the International Bureau (PCT Rule 17.2(a)). | | | | | | | | |
| * See the attached detailed Office action for a list of the certified copies not received. | | | | | | | | |
| | | | : | | | | | |
| Attachmen | t(s) | | | | | | | |
| | e of References Cited (PTO-892) | 4) Interview Summary | | • | | | | |
| 2) Notice of Draftsperson's Patent Drawing Review (PTO-948) 3) Information Disclosure Statement(s) (PTO/SB/08) Paper No(s)/Mail Date 5) Notice of Informal Patent Application | | | | | | | | |
| Paper No(s)/Mail Date 6) Other: | | | | | | | | |

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DETAILED ACTION

- 1. Claims 1 and 5-15 are pending.
- 2. In view of the amendment filed 8/11/06, the following rejections remain.
- 3. The following is a quotation of the appropriate paragraphs of 35 U.S.C. 103(a) that form the basis for the rejections under this section made in this Office Action:

A person shall be entitled to a patent unless:

- (a) A patent may not be obtained though the invention is not identically disclosed or described as set forth in section 102 of this title, if the differences between the subject matter sought to be patented and the prior art are such that the subject matter as a whole would have been obvious at the time the invention was made to a person having ordinary skill in the art to which said subject matter pertains. Patentability shall not be negatived by the manner in which the invention was made.
- 4. This application currently names joint inventors. In considering Patentability of the claims under 35 U.S.C. 103(a), the examiner presumes that the subject matter of the various claims was commonly owned at the time any inventions covered therein were made absent any evidence to the contrary. Applicant is advised of the obligation under 37 CFR 1.56 to point out the inventor and invention dates of each claim that was not commonly owned at the time a later invention was made in order for the examiner to consider the applicability of 35 U.S.C. 103(c) and potential 35 U.S.C. 102(e), (f) or (g) prior art under 35 U.S.C. 103(a).
- 5. Claims 1, and 11-15 stand rejected under 35 U.S.C. 103(a) as being unpatentable over US Pat No 5,753,228 (May 1998, PTO 892) or US Pat No 4,357,272 (Nov 1982, PTO 892) each in view of US Pat No 5,688,682 (Nov 1997; PTO 892) and Beasley *et al* (Food and Agricultural Immunology 12: 303-215, Sept 2000, PTO 892).

The '228 patent teaches a process for the production of egg yolk antibodies binding to any parasitic antigen wherein the reference method comprises the steps of selecting suitable poultry bird such as hens or chicken (See column 5, lines 6-9, in particular), immunizing the poultry birds such as the chicken with known complete adjuvant such as Freund's complete adjuvant containing heated killed and dried 1 mg/ml of M tuberculosis (See column 5, lines 13-21, Example 1, in particular). The '228 patent teaches that the adjuvant enhances the antibody responsiveness to the immunogen (See column 5, lines 13-18, in particular). The '228 patent teaches the bird such as leghorn hens, 21 weeks old are immunized with immunogen such as C

parvum in Freund's complete adjuvant and booster shots are given at intervals of five weeks (See column 8, lines 26-33, in particular). The reference method wherein the eggs are collected, stored at 4 °C until processed. The '228 patent teaches that the advantages of antibodies from egg yolks of hyperimmunized hens are: it provides a continuous source of large quantities of uniform antibodies which can be easily collected and stored (column 1, lines 54-59, in particular) and whole chicken serum is also remarkably resistant to temperature and acidity (See column 5, line 49-51, in particular).

The '272 patent teaches a process for the production of egg yolk antibodies binding to any antigen wherein the reference method comprises the steps of selecting suitable poultry bird such as Leghorn chicken (See column 6, Example 1, Hens, in particular), immunizing the poultry birds such as the chicken with any antigen in the range of 1 to 5 mg/ml which is equivalent to 1000 to 5000 µg/ml intramuscularly in known incomplete adjuvant (See column 6, lines Immunization, lines 40-55, in particular). The '272 patent teaches the concentration of antigen used is not critical and varied from one antigen to another, but is generally in the range of 1 to 5 mg/ml. After the initial injection, the hens are immunized with additional injections (booster shot) at weekly intervals until the state of hyperimmunization is reached. The hyperimmunized eggs are collected and stores at 4 °C until use and this continuous over a period of 9 months (See column 6, lines 47-54, in particular). The '272 patent further teaches that if the antigen has low molecular weight (non-immunogenic), the immunogenicity of antigen can be enhance by crosslinking with carbodimines (see column 5, lines 19-31, in particular). The '272 patent teaches that the antibody IgY titer produced ranges from 128 to 512 which is within the claimed range of 165-225 (see column 8, lines 6-10, in particular). The reference production of antibody is detectable 10 days after initial immunization and continued for 4 months (See column 7, lines 67 bridging column 8, lines 1-4, in particular). The '272 patent teaches the advantages of producing egg yolk antibodies are that it is comparatively easy to raise and keep chickens under conditions where they will produce antibody against the antigen desired and the antibody produced persists over such long periods such as the entire laying period (see column 4, lines 45-50, in particular).

The claimed invention in claim 1 differs from the teachings of the reference only in that the process for producing egg yolk antibodies binding to small molecule organo chlorine pesticides instead of parasitic antigens as taught by the '228 patent or any antigen as taught by the '272 patent wherein the bird is immunized with 1000 μg conjugate 2,4,5 trichlorphenoxyacetic acid β-alanine mixed in 0.85 ml paraffin and 0.15 ml mannide monooleate in breast muscle, and

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the booster shots are given at a dose of 500 μ g conjugate 2,4,5 trichlorphenoxyacetic acid β alanine at the intervals of two, three five weeks and five weeks thereof as long as the bird lays
eggs.

The claimed invention in claim 2 differs from the teachings of the references only in that the process for the production of egg yolk antibodies binding to small molecule organo chlorine pesticides wherein the desired hapten-protein conjugates having the binding properties to Hexachlorohexane.

The '682 patent teaches various adjuvant such as known complete adjuvant for vaccine Emulsigen, which is a paraffin oil in water emulsion that can be used in food animal and Freund's Incomplete adjuvant which is 15 percent (0.15 ml) by weight mannide monooleate and 85% (0.85 ml) paraffin oil (See column 4, lines 24-31, in particular). The reference adjuvants are useful for slowly releasing the vaccine into the animal and potentiating the immune response (See column 4, lines 31-32, in particular).

Beasley et al teach a method of making polyclonal antibody that binds to various pesticides such as Hexachlorohexane (HCH) by immunizing the rabbit with 2,4,5trichlorophenoxyacetic acid (2,4,5-T) having a β-alanine spacer arm conjugated to KLH or Ova where the reference antibody having the binding property to Hexachlorohexane (HCH) (See page 207, Antibody production, in particular). Beasley et al further teach a method of making hapten conjugate to small molecule organo chlorine pesticide such as herbicide 2,4,5trichlorophenoxyacetic acid (2,4,5-T) having a β-alanine spacer arm by hydroxysuccinimide (See page 205, Materials and Methods, second full paragraph, page 206, structure Ib, in particular). The reference process of making conjugate hapten 2, 4,5-Trichloro phenoxy acetic acid (TCB) hapten binding to hexachloro hexane involves the steps of: (a) adding β -alanine spacer arm to 2.55 g of 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) in 5.95 ml thionyl chloride (50 mmol), (b) refluxing for 1 hour and removing unreacted thionyl chloride by evaporation; (c) stirring the product with β-alanine (9 mmol, 0.66g in 7.4 ml of 1M OH) at 0°C; (d) then warming the product over 16 hours at room temperature; (e) isolating the resulting acid by acidification; (f) partitioning the into ethyl acetate; (g) washing with water and brine and giving a yield of 0.5g or 16% of crude product hapten containing 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) (See page 205, second full paragraph, in particular). Beasley et al teach 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) is slightly more sensitive than other Hapten protein conjugate such as 1,2,4-TCB (See page 209, line 1, in particular) and 2,4,5 trichlorophoxyacetic acid (2,4,5-TCP) provides a relative

simple targets for antibody development, enabling the detection of HCH residues by immunoassays after its conversion in samples to chlorobenzenes (See page 204, last paragraph, in particular).

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to substitute the parasitic immunogen as taught by the '228 patent or the antigen as taught by the '272 patent for the small organo chlorine pesticide immunogen such as 2,4,5-trichlorophenoxyacetic acid (2,4,5-T)-β-alaine as taught by Beasley *et al* in adjuvant such as 15 percent (0.15 ml) by weight mannide monooleate and 85% (0.85 ml) paraffin oil as taught by the '682 patent for a process of making any egg yolk antibodies that bind to small molecule organo chlorine pesticide as taught by the '228 patent, the 682 patent and Beasley *et al*. From the combined teachings of the references, it is apparent that one of ordinary skill in the art would have had a reasonable expectation of success in producing the claimed invention.

One having ordinary skill in the art would have been motivated to do this because the '228 patent teaches that the advantages of antibodies from egg yolks of hyperimmunized hens because it provides a continuous source of large quantities of uniform antibodies which can be easily collected and stored (column 1, lines 54-59, in particular) and whole chicken serum is also remarkably resistant to temperature and acidity (See column 5, line 49-51, in particular). The '272 patent teaches the advantages of producing egg yolk antibodies are that it is comparatively easy to raise and keep chickens under conditions where they will produce antibody against the antigen desired and the antibody produced persists over such long periods such as the entire laying period (see column 4, lines 45-50, in particular). The Beasley et al teach 2.4,5trichlorophenoxyacetic acid (2,4,5-T) is useful for making antibody that binds to Hexachlorohexane (HCH) and 2,4,5-T is slightly more sensitive than other Hapten protein conjugate such as 1,2,4-TCB (See page 209, line 1, in particular). The '682 patent teaches that adjuvant is useful for slowly releasing the vaccine into the animal and in potentiating the immune response (See column 4, lines 31-32, in particular). The booster shots of immunizing the bird again and again at various intervals such as two, three or five weeks as long as the bird lay eggs is within the purview of one skill in the art at the time the invention was made because the '228 patent and the '272 patent teach booster shots maintain hyperimmunized antibody producing state and enhance the titer of the antibody. The recitation of collecting the eggs daily and stored at 4 °C until use is within the purview of one ordinary skill in the art at the time the invention was made because it is routine and customary to store away the collected egg at room temperature or

refrigerated at 4°C until use. Claim 14 is included in this rejection because the sensitivity of the egg yolk antibody (polyclonal) is an inherent property of the antibody and would expect to be equally sensitive to the polyclonal or monoclonal antibodies produced by mammals because they are immunized with the same immunogen.

Applicants' arguments filed 8/11/06 have been fully considered but are not found persuasive.

Applicants' position is that the US Pat No. 5,753,228 teaches a process for the production of egg yolk antibodies binding to any parasitic antigen (i.e antigen from living organism) where as the current claim relates to antibodies against the organo chlorine chemicals. Similarly the present invention differs from the teachings of the prior art, as the present invention relates to the hapten protein conjugates and not about adjuvants. As it is understood that adjuvants are useful for the slow release of the vaccine whereas the haptens are high molecular weight molecules which aid the antigen being quickly recognized by the immune system. Beasley relates to the production of polyclonal antibody against *HCH* by immunizing rabbits, a type of mammal, whereas the pending claim relates to avians. It is accepted by those of skill in the art that every genus of animal has a unique immune system. The present invention also has a clear advantage over Beasley et al as the production rate and effectiveness of the antibodies from the host is very high when compared to the rabbits as used by Beasley at al.

In response, as stated by applicants, the US Pat No. 5,753,228 teaches a process for the production of egg yolk antibodies to parasitic antigen instead of organo chlorine chemicals whereas the claims are drawn to a process for the production of egg yolk antibodies to organo chlorine chemicals. However, Beasley *et al* teach a process of making hapten protein conjugated 2,4,5 tricholorophenoxyacetic acid β-alanine to be used as antigen for making antibody. Given the antigen is known as taught by Beasley et al and the process of making egg yolk antibodies as well as the advantage of making egg antibodies as taught by the '228 patent would have lead to one of ordinary skilled in the art at the time the invention was made to substitute the parasitic antigen in the process of producing egg yolk antibodies as taught by the '228 patent for the hapten protein conjugated 2,4,5 tricholorophenoxyacetic acid β-alanine antigen as taught by Beasley et al for making egg antibodies binding to small molecule organo chlorine pesticides. Further, the binding specificity of the claimed egg yolk as recited in claim 1 is to any small molecule organo chlorine pesticides in the claimed process, which encompasses the egg antibodies binding to hexachloro hexane (HCH) as taught by Beasley et al and the '228 patent.

6. Claim 5 stands rejected under 35 U.S.C. 103(a) as being unpatentable over US Pat No 5,753,228 (May 1998, PTO 892) or US Pat No 4,357,272 (Nov 1982, PTO 892) each in view of US Pat No 5,688,682 (Nov 1997; PTO 892) and Beasley *et al* (Food and Agricultural Immunology 12: 303-215, Sept 2000; PTO 892) as applied to claims 1 and 11-15 mentioned above and further in view of McAdam et al (J Agric Food Chem 40: 1466-70, 1992; PTO 892).

The combined teachings of the '228 patent, the '272 patent, the '682 patent and Beasley et al have been discussed supra. Beasley et al further teach a method of making hapten conjugate to small molecule organo chlorine pesticide such as herbicide 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) which conjugated to a β -alanine spacer arm by hydroxysuccinimide (See page 205, Materials and Methods, second full paragraph, page 206, structure Ib, in particular). The reference process of making conjugate hapten 2, 4,5-Trichloro phenoxy acetic acid (TCB) hapten binding to hexachloro hexane involves the steps of: (a) adding β -alanine spacer arm to 2.55 g of 2.4.5-trichlorophenoxyacetic acid (2.4.5-T) in 5.95 ml thionyl chloride (50 mmol), (b) refluxing for 1 hour and removing unreacted thionyl chloride by evaporation; (c) stirring the product with β-alanine (9 mmol, 0.66g in 7.4 ml of 1M OH) at 0°C; (d) then warming the product over 16 hours at room temperature; (e) isolating the resulting acid by acidification; (f) partitioning the into ethyl acetate; (g) washing with water and brine and giving a yield of 0.5g or 16% of crude product hapten containing 2,4,5-trichlorophenoxyacetic acid (2,4,5-T) (See page 205, second full paragraph, in particular). Beasley et al teach the impurity such as un-react hapten can be isolate by chromatography on silica gel (see page 205, last paragraph, in particular) using various solvents such as chloroform (see page 205, last paragraph) and methanol of choice (See page 211, first paragraph, in particular). The ratio of chloroform and methanol such as 85:15 as eluent for thin layer chromatography is within the purview of one skill in the art at the time the invention was made because it is a routine optimization to separate the product from the contaminant. The spraying with 2% o-toludine in acetone for thin layer chromatography is within the purview of one skill in the art at the time the invention was made because it is a routine visualization in thin layer chromatography analysis. The Rf value of 0.45 and the melting rang of 169-70°C are inherent properties of the reference compound. Beasley et al further teach synthesizing NHS ester of 2,4,5-trichlorophenoxyacetic acid (2,4,5-T)-β-alaline using the same procedure as Triclopyr by dissolving in dichloromethane, adding N-hydroxysuccinimide and dicyclohexycarbodiimide as a coupling agent and stirring at room temperature and isolating the product using silica (See page 205, last paragraph, in particular). Beasley et al teach the most

effective solvent for hexachlorocyclexane are methanol, acetone and hexane:acetone (4:1) (See page 210, Detection of Residues in Soil and Water, in particular).

The claimed invention in claim 5 differs from the combined teachings of the references only in that the process for the production of egg yolk antibodies to small molecule organo chlorine pesticides wherein the production of conjugate hapten 2,4,5-Trichloro phenoxy acetic acid β -alanine (TCB) hapten binding to Hexachloro hexane involves the steps of adding dimethylaminopyridine as a catalyst; stirring the mixture overnight and the temperature slowly raised to the room temperature; filtering and evaporating acetone and (r) separating the active ester as a colorless solid.

McAdam *et al* teach three approaches to hapten-protein conjugation such as coupling of β-alanine to organophosphate by dissolving β-alanine in dichloromethane, adding dicyclohexylcarbodiimide (DCC) and dimethylaminopyridine as a catalyst, stirring the mixture at room temperature such as 20°C, filtering, drying and separating the active ester (See page 1467, column 1, first paragraph, in particular). McAdam *et al* further teach a method of coupling of activated ester such as Fenitrothion to hapten such as KLH, HRP or Ovalbumin (OA) using N-hydroxysuccinimide as a coupling agent for making the reference ogranophosphate Fenitrothion more immunogenic for antibody production (See page 1467, column 1, Coupling of Activated Feitrothion Succinimide Esters to carrier proteins, column 2, polyclonal and monoclonal antibody production, in particular). McAdam *et al* teach that hapten conjugates coupled through the spacer-arm such as alanine yield the most specific monoclonal and polyclonal antibody and higher affinity (See page abstract, page 1468 column 2, last paragraph, in particular). McAdam *et al* teach Monoclonal antibodies offer the advantage of potential scale up of production of any well defined antibody and polyclonal antibodies prepared the same way used in the same assay format is only slightly less sensitive (See page 1469, column 2, General Discussion, in particular).

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to produce any egg yolk antibodies (polyclonal antibodies) binding to small molecule organochlorine pesticides as taught by the '228 patent, the '272 patent, the '682 patent and Beasley et al by synthesizing the active ester of any hapten-beta alanine by dissolving in dichloromethane, coupling to beta-alanine using N-hydroxysuccinimide and dicyclohexylcarbodiimide as coupling agents as taught by Beasley et al and McAdam et al in the present of dimethylaminopyridine as a catalyst as taught by McAdam et al. From the combined

teachings of the references, it is apparent that one of ordinary skill in the art would have had a reasonable expectation of success in producing the claimed invention.

One having ordinary skill in the art would have been motivated to do this because McAdam *et al* teach that hapten conjugates coupled through the spacer-arm such as alanine yield the most specific monoclonal and polyclonal antibody and higher affinity (See page abstract, page 1468 column 2, last paragraph, in particular). McAdam *et al* teach Monoclonal antibodies offer the advantage of potential scale up of production of any well defined antibody and polyclonal antibodies prepared the same way used in the same assay format is only slightly less sensitive (See page 1469, column 2, General Discussion, in particular). The recitation of adding dimethylsulphoxide (DMSO) drop wise to the mixture until the hapten dissolved is within the purview of one ordinary skill in the art at the time the invention was made because polar solvent such as DMSO dissolves similar polar compound (like dissolves like) since Beasley et al teach the most effective solvent for hexachlorocyclexane are methanol, acetone and hexane:acetone (4:1) (See page 210, Detection of Residues in Soil and Water, in particular). The recitation of melting range of 102-104 °C of 2,4,5-trichlorophenoxyacetic-beta alanine is inherent property of the compound 2,4,5-trichlorophenoxyacetic-beta alanine.

Applicants' arguments filed 8/11/06 have been fully considered but are not found persuasive.

Applicants' position is that the US Pat No. 5,753,228 teaches a process for the production of egg yolk antibodies binding to any parasitic antigen (i.e antigen from living organism) where as the current claim relates to antibodies against the organo chlorine chemicals. Similarly the present invention differs from the teachings of the prior art, as the present invention relates to the hapten protein conjugates and not about adjuvants. As it is understood that adjuvants are useful for the slow release of the vaccine whereas the haptens are high molecular weight molecules which aid the antigen being quickly recognized by the immune system. Beasley relates to the production of polyclonal antibody against *HCH* by immunizing rabbits, a type of mammal, whereas the pending claim relates to avians. It is accepted by those of skill in the art that every genus of animal has a unique immune system. The present invention also has a clear advantage over Beasley et al as the production rate and effectiveness of the antibodies from the host is very high when compared to the rabbits as used by Beasley at al.

In response, as stated by applicants, the US Pat No. 5,753,228 teaches a process for the production of egg yolk antibodies to parasitic antigen instead of organo chlorine chemicals

whereas the claims are drawn to a process for the production of egg yolk antibodies to organo chlorine chemicals. However, Beasley *et al* teach a process of making hapten protein conjugated 2,4,5 tricholorophenoxyacetic acid β -alanine to be used as antigen for making antibody. Given the antigen is known as taught by Beasley et al and the process of making egg yolk antibodies as well as the advantage of making egg antibodies as taught by the '228 patent would have lead to one of ordinary skilled in the art at the time the invention was made to substitute the parasitic antigen in the process of producing egg yolk antibodies as taught by the '228 patent for the hapten protein conjugated 2,4,5 tricholorophenoxyacetic acid β -alanine antigen as taught by Beasley et al for making egg antibodies binding to small molecule organo chlorine pesticides. Further, the binding specificity of the claimed egg yolk as recited in claim 1 is to any small molecule organo chlorine pesticides in the claimed process, which encompasses the egg antibodies binding to hexachloro hexane (HCH) as taught by Beasley et al and the '228 patent.

7. Claims 6 and 8-9 stand rejected under 35 U.S.C. 103(a) as being unpatentable over US Pat No 5,753,228 (May 1998, PTO 892) or US Pat No 4,357,272 (Nov 1982, PTO 892) each in view of US Pat No 5,688,682 (Nov 1997; PTO 892) and Beasley *et al* (Food and Agricultural Immunology 12: 303-215, Sept 2000; PTO 892) as applied to claims 1 and 11-15 mentioned above and further in view of Deignan *et al* (Food and Agricultural Immunology 12: 77-85, March 2000; PTO 892) and Akita *et al* (J of Food Science 57(3): 629-634, 1992; PTO 892).

The combined teachings of the '228 patent, the '272 patent, the '682 patent and Beasley et al have been discussed supra.

The claimed invention in claim 6 differs from the combined teachings of the references only that the process for the production of egg yolk antibodies wherein harvesting of antibodies as defined in step (g) of claim 1 (a) obtained egg yolk without rupturing the yolk; (b) adding 100 ml of Tris buffer for every 10 ml of yolk; (c) removing the precipitate by centrifugation; (d) adding to the supernatant the precipitate solution of magnesium chloride and phosphotungstic acid for centrifuging; (e) discarding the pellet; (f) adding to the supernatant a water solution protein fraction of 12% polyethylene glycol; (g) incubating for 10 minutes and then centrifuging again; (h) precipitating out the antibody; (i) adding 10 ml of 10mM phosphate buffer to dissolve the precipitate; (j) cooling the antibody solution to 0°C; (k) adding 10 ml of precooled ethanol; (l) centrifuging the solution at 4°C and dissolving the sediment in 10 mM phosphate buffer; and (m) dialyzing against phosphate buffer for 24 hour at 4°C to obtain the yield of antibodies.

The claimed invention in claim 8 differs from the combined teachings of the references only in that the process for the production of egg yolk antibodies to small organo chlorine pesticides wherein the lipid from egg yolk is precipitated out twice using the precipitating solution of phosphotungstic acid and magnesium chloride and centrifuged obtaining the antibody yield up to 75% from supernatant.

The claimed invention in claim 9 differs from the combined teachings of the references only in that the process for the production of egg yolk antibodies to small organo chlorine pesticides wherein the pH of the water soluble protein fraction obtained after the removal of the lipids is adjusted to pH 5.0 to further precipitate out the antibodies for obtaining a yield of 80-90%.

Deignan et al teach a comparative analysis of five published methods of purifying egg yolk immunoglobulin such as lipid removal by freeze and thaw at neutral pH of Jensenius et al (1981), precipitation with 3.5% polyethylene glycol (PEG) of Polson and von Wechmar (1980), precipitation with dextran sulphate and calcium chloride of Jensenius et al (1981), precipitation of with phosphotungstic acid and magnesium chloride of Vieria et al (1984) (See entire document, Lipid removal page 78 bridging page 79, in particular) and immunoglobulin precipitation by precipitation using 12% PEG of Polson & von Wedmar, 1980; Polson et al 1985) (See page 80, in particular). Deignan et al teach obtaining egg yolk without rupturing the yolk (page 78, egg yolk separation, in particular), follows by lipid removal using the method of Vieira et al by adding 100 ml of Tris buffer for every 10 ml of yolk; (c) removing the precipitate by centrifugation; (d) adding to the supernatant the precipitate solution of magnesium chloride and phosphotungstic acid for centrifuging; (e) discarding the pellet (See page 79, Precipitation with phosphotungstic acid and magnesium chloride, in particular), follows by immunoglobulin precipitation using the method of Polson & von Wedmar et al by adding to the supernatant a water solution protein fraction of 12% polyethylene glycol; (g) incubating for 10 minutes and then centrifuging again; (h) precipitating out the antibody; (i) adding 5 ml of 10mM phosphate buffer to dissolve the precipitate; (j) cooling the antibody solution to 0°C; (k) adding 5 ml of precooled ethanol; (l) centrifuging the solution at 4°C and dissolving the sediment in 5ml of phosphate buffer; and (m) dialyzing against phosphate buffer for 24 hour at 4°C to obtain the yield of antibodies. Deignan et al teach that the advantage of removal of lipid from native egg yolk using a combination of polyanions and cations such as phosphotungstic acid and magnesium chloride it that it recovered the highest yield of 21.6 mg (range of 20.4-33.0) of protein per ml of egg yolk (See Figure 1,

page 81, in particular) and the IgY purity as estimated by densitometry was 69.8%. Following lipid removal, immunoglobulin precipitation using 12% PEG gives the highest yield of 8.62 mg (range 8.39 to 8.83) or IgY per ml of egg yolk and this method was deemed the best (See page 82, Ig precipitation, Fig 2, page 82, Discussion, in particular).

Akita *et al* teach a process of purifying egg antibodies (IgY) by lowering the pH to 5.0 of the water soluble protein fraction (WSF) to further remove the lipid from said WSF and the highest yield of IgY such as 92.7 to 94.2 % is obtained between 5.0 to 5.2, respectively (See Table 1, page 631, in particular).

Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to harvest the egg yolk antibodies that bind to small molecule organo chlorine pesticides as taught by the '228 patent, the '272 patent, the '682 patent and Beasley et al using the solution of phosphotungstic acid and magnesium chloride follows by Ig precipitation using 12% PEG as taught by the Deignan et al and further remove the lipid from said WSF by lowering the pH to 5.0 as taught by Akita et al. From the combined teachings of the references, it is apparent that one of ordinary skill in the art would have had a reasonable expectation of success in producing the claimed invention.

One having ordinary skill in the art would have been motivated to do this because Deignan *et al* teach that the advantage of removing lipid from native egg yolk using a combination of polyanions and cations such as phosphotungstic acid and magnesium chloride is that it recovered the highest yield of 21.6 mg (range of 20.4-33.0) of protein per ml of egg yolk (See Figure 1, page 81, in particular) and the IgY purity as estimated by densitometry was 69.8%. Following lipid removal, immunoglobulin precipitation using 12% PEG gives the highest yield of 8.62 mg (range 8.39 to 8.83) or IgY per ml of egg yolk and this method is deemed the best (See page 82, Ig precipitation, Fig 2, page 82, Discussion, in particular). The recitation of precipitated out lipid from egg yolk "twice" using the precipitating solution of phosphotungstic acid and magnesium chloride is within the purview of one ordinary skill in the art at the time the invention was made because it is an obvious variation of the teaching of Deignan *et al* who teaches that the advantage of removal of lipid from native egg yolk using a combination of polyanions and cations such as phosphotungstic acid and magnesium chloride would give the IgY purity as estimated by densitometry was 69.8%, which is within the limit of "up to" 75%". Akita *et al* teach that the advantage of lowering the pH to 5.0 of the water soluble protein fraction (WSF)

further removes the lipid from said WSF and the highest yield of IgY such as 92.7 to 94.2 % is obtained between 5.0 to 5.2, respectively (See Table 1, page 631, in particular).

Applicants' arguments filed 8/11/06 have been fully considered but are not found persuasive. Applicants further wish to emphasize that the process of the production of the egg yolk antibodies defined briefly in the claim 6 differs from the teachings of the prior art, as the harvesting of the same is carried out in the process not taught in the prior art. Moreover, in the claimed invention, claim 8 differs from the combined teachings of the reference, that the process of production of egg yolk antibodies to small organo chlorine pesticides where in the lipid content from the egg yolk is precipitated twice to yield 75% from supernatant against the teaching of prior art. Claim 9 further differs from the combined references, that the process for the production of egg yolk antibodies to small organo chlorine compounds by haptens, whereas the prior art deals with the antibody production against the parasites or parasitic antigens from rabbits.

In response to the argument that the claimed process for the production of egg yolk antibodies to small organo chlorine compounds by haptens, whereas the prior art deals with the antibody production against the parasites or parasitic antigens from rabbits, once a prima facie case of obviousness has been made the burden of going further is shifted to applicant. In re Keller, 642 F.2d 4B, 208 USPQ 871, 882 (CCPA 1981). This applicant has not done, but rather argues the references individually and not their combination. One cannot show non-obviousness by attacking references individually where the rejections are based on a combination of references. In re Young 403 F.2d 759, 150 USPQ 725 (CCPA 1968).

In response to the argument that the process of the production of the egg yolk antibodies defined briefly in the claim 6 differs from the teachings of the prior art, Deignan *et al* teach obtaining egg yolk without rupturing the yolk (page 78, egg yolk separation, in particular), follows by lipid removal using the method of Vieira et al by adding 100 ml of Tris buffer for every 10 ml of yolk; (c) removing the precipitate by centrifugation; (d) adding to the supernatant the precipitate solution of magnesium chloride and phosphotungstic acid for centrifuging; (e) discarding the pellet (See page 79, Precipitation with phosphotungstic acid and magnesium chloride, in particular), follows by immunoglobulin precipitation using the method of Polson & von Wedmar et al by adding to the supernatant a water solution protein fraction of 12% polyethylene glycol; (g) incubating for 10 minutes and then centrifuging again; (h) precipitating out the antibody; (i) adding 5 ml of 10mM phosphate buffer to dissolve the precipitate; (j) cooling

the antibody solution to 0°C; (k) adding 5 ml of precooled ethanol; (l) centrifuging the solution at 4°C and dissolving the sediment in 5ml of phosphate buffer; and (m) dialyzing against phosphate buffer for 24 hour at 4°C to obtain the yield of antibodies. It is within the purview of one of ordinary skill in the purification art to add sufficient phosphate buffer to dissolve the antibody precipitates. The recitation of 10 ml instead of 5 ml is an obvious variation of the reference teachings.

In response to the argument that claim 8 differs from the combined teachings of the reference since the process of production of egg yolk antibodies to small organo chlorine pesticides where in the lipid content from the egg yolk is precipitated twice to yield 75% from supernatant against the teaching of prior art, claim 8 recites the process of producing egg antibodies wherein the egg yolk is precipitated out twice using the phosphotungstic acid and magnesium chloride and then centrifuged to yield antibody up to 75% from the supernatant. Claim 8 does not recite the process of production of egg yolk antibodies to small organo chlorine pesticides where in the lipid content from the egg yolk is precipitated twice to yield 75% from supernatant as argued. The term "up to 75%" could be any where from 69.8% to 75%, which is within the limit of "up to" 75%". Deignan et al teach the egg yolk is precipitated out using the phosphotungstic acid and magnesium chloride by adding to the supernatant the precipitate solution of magnesium chloride and phosphotungstic acid for centrifuging; (e) discarding the pellet (See page 79, Precipitation with phosphotungstic acid and magnesium chloride, in particular), follows by immunoglobulin precipitation using the method of Polson & von Wedmar et al by adding to the supernatant a water solution protein fraction of 12% polyethylene glycol; (g) incubating for 10 minutes and then centrifuging again; (h) precipitating out the antibody. The number of time such as twice using phosphotungstic acid and magnesium chloride to precipitate out egg antibodies is within the purview of one of ordinary skill in the antibody purification to adjust the number of precipitation using the same solution as taught by Deignan et al.

8. Claims 7 and 10 are rejected under 35 U.S.C. 103(a) as being unpatentable over US Pat No 5,753,228 (May 1998, PTO 892) or US Pat No 4,357,272 (Nov 1982, PTO 892) each in view of US Pat No 5,688,682 (Nov 1997; PTO 892) and Beasley *et al* (Food and Agricultural Immunology 12: 303-215, Sept 2000; PTO 892) as applied to claims 1 and 11-15 mentioned above and further in view of Akita *et al* (J of Immunological Methods 160: 207-214, 1993; PTO 892) or Hatta *et al* (Agric Biol Chem 54(10): 2531-2535, 1990; PTO 892).

The combined teachings of the '228 patent, the '272 patent, the '682 patent and Beasley et al have been discussed supra.

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The claimed invention in claim 7 differs from the combined teachings of the references only in that the process for the production of egg yolk antibodies to small organo chlorine pesticides wherein the harvesting of antibodies can also be conducted as follows: (a) obtaining the egg yolk from the eggshell without rupturing the yolk membrane; (b) adding for every 10 ml of yolk, 10 ml of distilled water; (c) adding about 0.15% of kappa-carrageenan and left to stir for 30 minutes at room temperature; (d) filtering and centrifuging the solution for 15 minutes; (e) passing through the DEAE-Sephacel column prepared with 20 mM phosphate buffer at pH 8.0; (f) eluting with 0.2 M phosphate buffer pH 8.0; (g) collecting the eluate and the absorbance read at 280 nm; and (h) pooling and storing the peak fractions containing the antibody at 4°C.

The claimed invention in claim 10 differs from the combined teachings of the references only in that the process for the production of egg yolk antibodies to small organo chlorine pesticides wherein the yield of antibody is to the extent of 73%.

Akita *et al* teach a process of isolating egg yolk immunoglobulin such as obtaining egg yolk from the egg shell without rupturing the yolk membrane from immunized hens; adding about 0.15% (w/v) of carrageenan (120 mg in 80 ml distilled water, which equal to 0.15%) and mixing and stirring for 15 minutes at room temperature which is about 20° C, centrifuging it to separate the water-soluble protein faction from the yolk lipoproteins (see page 210, right column, Fig 4, in particular). Akita *et al* teach the reference method yield 89% or 7.3 mg/ml of egg yolk (See caption in Figure 4, in particular). Akita *et al* teach the IgY in water-soluble protein fraction is purified by gel filtration such as passing through Sephadex G-25 column with the appropriate buffer and collecting the eluate by monitoring the absorbance at 280 nm (See page 210, Gel filtration, in particular). Akita *et al* further teach the hyperimmunized eggs are collected and store at 4°C until use (See page 208, column 2, immunization, in particular).

Hatta et al teach a process of isolating egg yolk immunoglobulin, IgG, a livetin protein, using several natural gums such as carrageenan and xanthan gum) by (a) obtaining the egg yolk from the eggshell without rupturing the yolk membrane from immunized hens; (b) adding about 0.1% (w/v) of kappa carrageenan (60 mg of carrageenan in 40 ml of distilled water) to the egg yolk in order to separate the water-soluble protein faction from the yolk lipoproteins by centrifugation (See page 2534, Diagram 1, Table 1, in particular); filtering the water-soluble protein fraction through filter paper, and passing through the DEAE-Sephacel column prepared

with 20 mM phosphate buffer at pH 8.0 (see page 2533, column 1, Purification of IgY from egg yolk, in particular); eluting the yolk antibodies with 0.2 M (200mM) phosphate buffer pH 8.0; collecting the peak fractions by monitoring at 280 nm (See page 2534, Diagram 1, Table 1, in particular). Hatta *et al* teach the purity of IgY obtained by the reference method was 98.3% with a yield of 73% (See page 2534, column 1, second full paragraph, in particular). Hatta *et al* teach natural gums such as kappa carrageenan are effective as precipitant of yolk lipoproteins and the gum has been used as a food ingredient, so that IgY prepared by this method should be suitable for oral administration (See page 2534, column 2, Discussion, in particular).

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Therefore, it would have been obvious to one of ordinary skill in the art at the time the invention was made to harvest the egg yolk antibodies that bind to small molecule organo chlorine pesticides as taught by the '228 patent, the '272 patent the '682 patent and Beasley et al using the process of separating lipoprotein by carrageenan and gel filtration such as DEAE-Sephacel column chromatography as taught by Akita et al and Hatta et al. From the combined teachings of the references, it is apparent that one of ordinary skill in the art would have had a reasonable expectation of success in producing the claimed invention.

One having ordinary skill in the art would have been motivated to do this because Akita et al teach that purification of IgY from egg yolk by carrageenan has no adverse effect on the immunoreactivities of IgY and the yield in terms of purity is 89% (See page 210, caption in Fig 4, abstract, in particular). Hatta *et al* teach natural gums such as kappa carrageenan are effective as precipitant of yolk lipoproteins and the gum has been used as a food ingredient, so that IgY prepared by this method should be suitable for oral administration (See page 2534, column 2, Discussion, in particular).

Applicants' arguments filed 8/11/06 have been fully considered but are not found persuasive.

Applicants' position is that the US Pat No. 5,753,228 teaches a process for the production of egg yolk antibodies binding to any parasitic antigen (i.e antigen from living organism) where as the current claim relates to antibodies against the organo chlorine chemicals. Similarly the present invention differs from the teachings of the prior art, as the present invention relates to the hapten protein conjugates and not about adjuvants. As it is understood that adjuvants are useful for the slow release of the vaccine whereas the haptens are high molecular weight molecules which aid the antigen being quickly recognized by the immune system. Beasley relates to the production of polyclonal antibody against *HCH* by immunizing rabbits, a type of mammal,

whereas the pending claim relates to avians. It is accepted by those of skill in the art that every genus of animal has a unique immune system. The present invention also has a clear advantage over Beasley et al as the production rate and effectiveness of the antibodies from the host is very high when compared to the rabbits as used by Beasley at al.

In response, as stated by applicants, the US Pat No. 5,753,228 teaches a process for the production of egg yolk antibodies to parasitic antigen instead of organo chlorine chemicals whereas the claims are drawn to a process for the production of egg yolk antibodies to organo chlorine chemicals. However, Beasley *et al* teach a process of making hapten protein conjugated 2,4,5 tricholorophenoxyacetic acid β-alanine to be used as antigen for making antibody. Given the antigen is known as taught by Beasley et al and the process of making egg yolk antibodies as well as the advantage of making egg antibodies as taught by the '228 patent would have lead to one of ordinary skilled in the art at the time the invention was made to substitute the parasitic antigen in the process of producing egg yolk antibodies as taught by the '228 patent for the hapten protein conjugated 2,4,5 tricholorophenoxyacetic acid β-alanine antigen as taught by Beasley et al for making egg antibodies binding to small molecule organo chlorine pesticides. Further, the binding specificity of the claimed egg yolk as recited in claim 1 is to any small molecule organo chlorine pesticides in the claimed process, which encompasses the egg antibodies binding to hexachloro hexane (HCH) as taught by Beasley et al and the '228 patent.

9. No claim is allowed.

10. THIS ACTION IS MADE FINAL. See MPEP § 706.07(a).

Applicant is reminded of the extension of time policy as set forth in 37 CFR 1.136(a). A shortened statutory period for response to this final action is set to expire THREE MONTHS from the date of this action. In the event a first response is filed within TWO MONTHS of the mailing date of this final action and the advisory action is not mailed until after the end of the THREE-MONTH shortened statutory period, then the shortened statutory period will expire on the date the advisory action is mailed, and any extension fee pursuant to 37 CFR 1.136(a) will be calculated from the mailing date of the advisory action. In no event will the statutory period for response expire later than SIX MONTHS from the date of this final action.

11. Any inquiry concerning this communication or earlier communications from the examiner should be directed to Phuong Huynh "NEON" whose telephone number is (571) 272-0846. The examiner can normally be reached Monday through Friday from 9:00 am to 5:30 p.m. A message may be left on the examiner's voice mail service. If attempts to reach the examiner by telephone are unsuccessful, the examiner's supervisor, Christina Chan can be reached on (571) 272-0841. The IFW official Fax number is (571) 273-8300.

12. Any information regarding the status of an application may be obtained from the Patent Application Information Retrieval (PAIR) system. Status information for published applications may be obtained from either Private PAIR or Public PAIR. Status information for unpublished applications is available through Private PAIR only. For more information about the PAIR system, see http://pair-direct.uspto.gov. Should you have questions on access to the Private PAIR system, contact the Electronic Business Center (EBC) at 866-217-9197 (toll-free).

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Patent Examiner

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